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PHENOLIC ACIDS AND FLAVONOIDS OF THE SPORE-BEARING

STEMS OF Equisetum arvense

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The spore-bearing stems of *Equisetum arvense* L. (field horsetail) develop earlier than the sterile stems (herbage) and die off after the formation of the spores. Freshly collected spore-bearing stems (Irkutsk region, end of May, moisture content 73-75%) were extracted with methanol, and the extract was treated successively with chloroform, diethyl ether, ethyl acetate, and butanol.

From the ether-soluble fraction by preparative chromatography on polyamide sorbent and elution with water and a mixture of water and ethanol we obtained phenolic acid fractions, and by elution with chloroform and chloroform methanol (with increasing proportions of methanol from 5 to 30%) fractions enriched with flavonoid compounds.

The phenolic acids and the flavonoids in the form of their TMS ethers were analyzed by GLC on the Tsvet-4 chromatograph with a flame-ionization detector under the conditions described previously [1, 2].

Acids and flavonoids were identified by the method of additives and by comparison of the retention times of the TMS ethers with those of authentic samples. Below we give the relative retention times (RRTs) of the TMS ethers of the phenolic acids.

TMS Derivatives of the Acids	RRT
p-Hydroxybenzoic	0.63
Vanillic*	1.00
Protocatachuic	1.25
p-Coumaric	1.85
Ferulic	3.08
Caffeic	3.78

*The retention time of the standard was 5 min.

In the flavonoid fraction we identified the following compounds:

TMS Derivatives of the Flavonoids	RRT
Naringenin*	1.03
Dihydrokaempferol	1.24
Dihydroquercetin	1.48
Apigenin	1.95
Luteolin	2.71
*The retention time of the standard was 7.25 min perature 288°C).	(column tem-

Irkutsk Institute of Organic Chemistry, Siberian Branch of the Academy of Sciences of the USSR. I. M. Sechenov First Moscow Medical Institute. Translated from Khimiya Prirodnykh Soedinenii, No. 6, pp. 803-804, November-December, 1978. Original article submitted June 30, 1978. The relative retention times of the phenolic acids and flavonoids investigated coincided with those of authentic samples. It must be mentioned that, of the phenolic acids, p-hydroxybenzoic, protocatachuic, and p-coumaric predominated quantitatively, while the others were present in only small amounts. Among the flavonoids, the flavones apigenin and luteolin predominated.

In contrast to the herbage of Equisetum arvense, no flavonols were detected in the sporebearing stems.

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FLAVONOIDS FROM Phlomis agraria

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Continuing a study of the composition of the flavonoids of Siberian species of Jerusalem sage, from the herbage of *Phlomis agraria* Bge. (family Labiatae) we have isolated three flav-onoids.

 $\frac{\text{Substance (I): composition C_2_1H_{18}O_{12}, mp 190-192°C; UV spectrum, nm: } \lambda_{\max}^{\text{init}} 350, 276,}{\lambda_{\max}^{+\text{CH}_3\text{COONa}} 352, 256; \lambda_{\max}^{+\text{C}_2\text{H}_5\text{ONa}} 410, 265 nm; \lambda_{\max}^{+\text{AlCl}_3} 390, 267; \lambda_{\max}^{+\text{H}_3\text{BO}_3+\text{CH}_3\text{COONa}} 372, 258.}$ In the products of enzymatic hydrolysis we detected β -glucuronic acid. On the basis of these facts substance (I) was identified as luteolin 7- β -glucuronide [1, 3].

Substance (II): had the composition $C_{21}H_{20}O_{11}$, mp 255°C. UV spectrum, nm: $\lambda_{\max}^{\text{init}}$ 249, 268, 255; $\lambda_{\max}^{+CH_3COONa}$ 352, 268, 257; $\lambda_{\max}^{+C_2H_3ONa}$ 404, 263; $\lambda_{\max}^{+A1Cl_3}$ 416, 278; $\lambda_{\max}^{+H_3BO_3+CH_3COONa}$ 374, 259. β -Glucose was detected in the products of enzymatic hydrolysis. These facts permitted substance (II) to be identified as luteolin 7- β -glucoside [2, 3].

Substance (III) is also a luteolin 0-glycoside, and its study is continuing.

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